# Histone H3K27ac (Acetyl Lys27) antibody - ChIP grade

# Cat. No. GTX60815

Host	Rabbit
Clonality	Polyclonal
lsotype	IgG
Application	WB, ICC/IF, Dot, ELISA, ChIP assay, ChIP-seq
Reactivity	Human, Mouse, Rat, Arabidopsis thaliana

Reference (3) Package 50 µg

## APPLICATION

### **Application Note**

\*Optimal dilutions/concentrations should be determined by the researcher.

Suggested dilution	Recommended dilution
WB	1:1,000
ICC/IF	1:500
Dot	1:20,000
ELISA	1:500
ChIP assay	0.5-5 µg
ChIP-seq	Assay dependent

Not tested in other applications.

PROPERTIES	
Form	Liquid
Buffer	PBS
Preservative	0.05% Sodium azide, 0.05% ProClin 300
Storage	Store as concentrated solution. Centrifuge briefly prior to opening vial. For short-term storage (1-2 weeks), store at 4°C. For long-term storage, aliquot and store at -20°C or below. Avoid multiple freeze-thaw cycles.
Concentration	2.8 mg/ml (Please refer to the vial label for the specific concentration.)
Immunogen	The region of histone H3 containing the acetylated lysine 27 (H3K27ac), using a KLH-conjugated synthetic peptide.
Purification	Purified by affinity chromatography
Conjugation	Unconjugated



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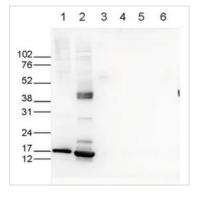


Note

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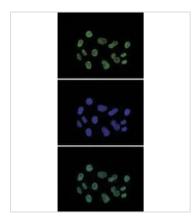
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DATA IMAGES



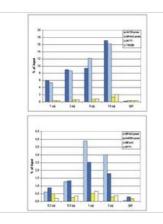
#### GTX60815 WB Image

WB analysis of whole cell (25 µg, lane 1) and histone extracts (15 µg, lane 2) from HeLa cells, and on 1 µg of recombinant histone H2A, H2B, H3 and H4 (lane 3, 4, 5 and 6, respectively) using GTX60815 Histone H3K27ac (Acetyl Lys27) antibody - ChIP grade. Dilution : 1:1,000



#### GTX60815 ICC/IF Image

ICC/IF analysis of 4% paraformaldehyde fixed HeLa cells using GTX60815 Histone H3K27ac (Acetyl Lys27) antibody - ChIP grade. Green : Primary antibody Blue : DAPI Dilution : 1:500



#### GTX60815 ChIP assay Image

ChIP analysis of sheared chromatin from  $10^6$  HeLa cells using GTX60815 Histone H3K27ac (Acetyl Lys27) antibody - ChIP grade. A titration consisting of 0.2, 0.5, 1 and 2 µg of antibody per ChIP experiment was analyzed. IgG (1 µg/IP) was used as a negative IP control. Quantitative PCR was performed with primers for the promoters of the active GAPDH and EIF4A2 genes, used as positive controls, and for the coding regions of the inactive MB and MYT1 genes, used as negative controls. This figure shows the recovery, expressed as a % of input (the relative amount of immunoprecipitated DNA compared to input DNA after qPCR analysis).



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